

ISOLATION AND CHARACTERIZATION OF BIOACTIVE COMPOUNDS FROM
SELECTED MANGROVE ENDOPHYTIC FUNGI ALONG THE KENYA COAST

HELEN MWAKA KITI

A THESIS SUBMITTED TO THE SCHOOL OF APPLIED AND HEALTH SCIENCES
IN THE DEPARTMENT OF PURE AND APPLIED SCIENCES IN PARTIAL
FULFILMENT OF THE REQUIREMENTS FOR THE AWARD OF DEGREE OF
DOCTOR OF PHILOSOPHY (CHEMISTRY) OF TECHNICAL UNIVERSITY OF
MOMBASA

2023

DECLARATION

Declaration by the Student

This thesis is my original work and has not been presented for examination in any University.

Helen Mwaka Kiti

PDC/0002/2016

Signature.....

Date.....

Declaration by the Supervisors

This thesis has been submitted with our approval as University supervisors.

Signature..... Date.....

Prof. Josiah Ochieng Odalo

Technical University of Mombasa

Sign.....Date.....

Prof. Paul Mwashimba Guyo

Pwani University

Sign.....Date.....

Dr. Cosmas Nzaka Munga

Technical University of Mombasa

Sign..... Date.....

Dr. Cromwell Mwiti Kibiti

Technical University of Mombasa,

DEDICATION

This thesis is dedicated to God Almighty and my family.

ACKNOWLEDGEMENT

Every departure must be accompanied by a destination in the far or near future. The long academic journey of my PhD has finally come to a good end after six years of hard work. First: I am grateful to the Technical University of Mombasa (TUM) for giving me the opportunity to study chemistry, and for granting me study leave. . Secondly, National Research Fund (NRF) Kenya for financial support through the post-graduate research grant NRF/R/2016/2017. Thirdly, Western Indian Ocean Marine Science Association (WIOMSA) for their financial support through the Marine Research Grant (MARG-1) Programme that supplemented my research budget. My sincere gratitude to Prof. Martin Onani, University of the Western Cape, South Africa and Prof. Matthias Heydenreich - Institut für Chemie, Postdam, Germany for all the nuclear magnetic resonance (NMR) analysis conducted in this study.

I trace my PhD journey to a conference sponsored by Kenya Coastal Development Project (KCDP) held at Voi Lodge, Taita Taveta County in March, 2016. This is where I met Dr. Elisha Mrabu of Kenya Marine and Fisheries Research Institute (KMFRI) and Dr. Joyce Jefwa of the National Museums of Kenya who inspired me to delve into the study of mangrove endophytic fungi. My sincere gratitude to these two for their inspiration. I would like to sincerely thank Prof. Josiah Ochieng Odalo, the main supervisor who has consistently and tirelessly provided support since the conceptualization of this project, through funding acquisition and to this end. I am truly grateful. I would wish to extend my gratitude to Prof. Paul Mwashimba Guyo, Pwani University for his guidance and support in providing valuable links during the PhD Programme. Thank you Dr. Cosmas Nzaka Munga for being a mentor. You have walked with me throughout proposal writing, funds acquisition, analysis and manuscript writing. Last but not least, my gratitude to Dr. Cromwell Kibiti for his guidance and support in thesis and manuscript writing. Thanks for always raising the red flag, it was elemental.

My sincere thanks to Dr. Mackenzie Nzaro, Chairperson in the Department of Pure and Applied Sciences and Dr. Udu Rahma, the former Dean School of Applied and Health Sciences for their great support. Laboratory technologists; Kennedy Agoi and Mercy Nyaribo of TUM, and Mackmillan Odhiambo of KMFRI for their support. I am grateful for all the support given to me by my colleagues in the Department of Pure and Applied Sciences.

I acknowledge and appreciate the support given to me by my family. First to my father, Mr. Simon Kiti Mbura for providing me and my siblings with equal opportunities for basic education. God bless you baba. My late mum, Jedilia Majala from whom I first learned that God answers the prayers of mortals and that victory belongs to everyone who believes. My husband Masai Mwawira for all your support. God bless your life. To our children for being a source of inspiration. Special thanks to Samuel Mazera, differently abled and for being the rate determining individual in all my undertakings; Esther Masai and Chao, always hopeful and cheerful, and Miriam Masai, though not understanding many things but conscious in many aspects that this study was taking a toll on me. Thank you, Miriam, for believing without understanding that somehow “mama” was doing the right things. For both the moral and financial support from my family, I am grateful. To my brother Samuel Jumaa, my sisters, Mrs. Ruth Wambua, Mrs. Claris Mbega, Mrs. Peninah Chesitoni for cheering me and believing in me. Thank you Rev. Silas Kazungu, Mrs. Lucy Mwandisha, Elizabeth Chacha, and Janet Kingi for your prayer support.

Finally, my uttermost gratitude goes to the Almighty God, the giver of life, the creator of the heavens and the earth. I am grateful for good health, the peace of mind, opening doors for funding, and causing the right people to come my way to either lead me, correct me or just encourage me along. I am forever grateful Lord. Thank you, Jesus.

TABLE OF CONTENTS

DECLARATION	ii
DEDICATION	III
ACKNOWLEDGEMENT.....	IV
TABLE OF CONTENTS.....	IV
LIST OF FIGURES.....	XII
LIST OF TABLES.....	VI
LIST OF ABBREVIATIONS.....	XX
ABSTRACT.....	XXIII
CHAPTER ONE	1
INTRODUCTION	1
1.1 Background Information	1
1. 2 Problem Statement.....	3
1.3 Justification of the Study	3
1.4 Significance of the Study	5
1.5 Objective.....	6
1.5.1 Overall Objective.....	6
1.5.2 Specific Objectives.....	6
1.6 Research Questions.....	7
1.7 Scope of the Study.....	7
CHAPTER TWO.....	8
LITERATURE REVIEW	8
2.1 Introduction	8
2.2 Infectious Diseases and Drug Resistance.....	8

2.2.1 Pathogenesis of <i>Staphylococcus Aureus</i>	10
2.2.1.1 Treatment and Control of <i>Staphylococcus Aureus</i>	10
2.2.2 Pathogenesis of <i>Escherichia coli</i>	11
Treatment and Control of <i>Escherichia coli</i>	12
2.3 Natural Bioactive Compounds.....	12
2.4 Sources of Natural Bioactive Compounds.....	14
2.4.1 Animal Sources of Natural Bioactive Products.....	14
2.4.2 Plants as Sources of Natural Bioactive Compounds.....	15
2.4.2.1 Alkaloids	16
2.4.2.2 Terpenoids	17
2.4.2.3 Saponins	18
2.4.2.4 Phenolics	18
2.4.2.5 Lipids.....	19
2.4.2.6 Polyketides	20
2.4.2.6.1 Biosynthesis of Beta Polyketides	21
2.4.2.7 Carbohydrates and Their Derivatives.....	23
2.4.2.8 Glycosides	24
2.4.3 Microorganisms as Sources of Natural Bioactive Compounds.....	24
2.4.3.1 Fungi	29
2.4.3.2 Endophytes	31
2.4.3.3 Endophytic Fungi	33
2.4.3.4 Fungi as Sources of Bioactive Compounds.....	34
2.4.3.5 Endophytic Fungi as Sources of Natural Bioactive Compounds	35
2.4.3.5.1 <i>Aspergillus as a Source of Natural Bioactive Compounds</i>	36
2.5 Mangroves as Reservoirs of Endophytic Fungi.....	37
CHAPTER THREE	42
MATERIALS AND METHODS.....	42
3.1 Introduction	42

3.2 Description of the Study Area	42
3.3 Preparation for Field and Laboratory Works	44
3.4 Experimental Procedures	44
3.5 Sampling Techniques and Illustrations	47
3.6 Ethical consideration.....	47
3.7 Sampling of Twigs from Selected Mangroves Species of Coastal Kenya	47
3.8 Sample Preparation.....	48
3.9 Isolation and Purification of Mangrove Fungal Endophytes.....	48
3.10 Morphological Diversity of Mangrove Fungal Endophytes	49
3.10.1 Cultural Identification of Fungal Isolates.....	50
3.10.2 Morphological Identification	50
3.11 Antimicrobial Properties of Mangrove Fungal Isolates	50
3.11.1 Small Scale Fermentation and Extraction of Fungal Metabolites.....	51
3.11.2 Preparation of Muller Hinton Agar Plates	52
3.11.3 <i>In vitro</i> Antimicrobial Susceptibility Test by Disk Diffusion Method....	53
3.11.4 Determination of Minimum Inhibition Concentration (MIC) Using Micro Dilution Method.....	55
3.12 Molecular Phylogenetic Characterization of the Active Fungal Endophytes	
3.12.1 Genomic DNA Extraction	56
3.12.2 Gel Electrophoresis.....	57
3.12.3 DNA Quantification	57
3.12.4 Polymerase Chain Reaction (PCR).....	58
3.12.5 Quality Control.....	58
3.13 Extraction, Purification, Isolation and Characterization of Secondary Metabolites of <i>Aspergillus flavus</i> of Coastal Kenya	58
3.13.1 Mass Culture and Extraction of Secondary Metabolites.....	59
3.13.2 Chemical Isolation and Purification of Secondary Metabolites <i>A. flavus</i>	59
3.13.2.1 Thin Layer Chromatography of the Crude Extracts	60

3.13.2.2 Silica Gel Column Chromatography.....	60
3.14 Data Analysis	61
3.14.1 Determination of Colonization Rate, Isolation Rate and Relative Frequency.....	61
3.14.2 Determination of the Diversity of Fungal Isolates.....	62
3.14.3 Antimicrobial Data Analysis.....	63
3.14.4 DNA Sequence, Editing and Phylogenetic Analyses of the Endophytes	63
3.14.5 Gas Chromatography-Mass Spectrometry (GC-MS) Analysis of Crude Ethyl Acetate Extracts <i>Aspergillus flavus</i> (MZ314728).....	64
3.14.6 Nuclear Magnetic Resonance (NMR) Analysis of Pure Compounds	65
CHAPTER FOUR.....	67
RESULTS	67
4.1 Introduction	67
4.1.1 Culturally Identified Mangrove Fungal Endophytes.....	67
4.1.2 Morphologically Identified Mangrove Fungal Endophytes.....	68
4.1.3 Colonization and Isolation Rate	74
4.2 Biological Characteristics of Mangrove Fungal Endophytes.....	76
4.2.1 Preliminary Screening of Mangrove Fungal Extracts.....	76
4.2.2 Minimum Inhibition Concentration (MIC) of Crude Extracts	81
4.3 Molecular Characteristics of Selected Mangrove Fungal Endophytes.....	82
4.4 Extracted, Purified and Characterized Secondary Metabolites from <i>Aspergillus Flavus</i>	87
4.4.1 Crude Extracts of <i>Aspergillus Flavus</i>	87
4.4.2 Constituents of Ethyl Acetate Crude Extracts of <i>Aspergillus flavus</i> by GC- MS Analysis	88
4.8. Results of GC-MS analysis of ethyl acetate crude extracts of <i>Aspergillus flavus</i> 4.4.3 TLC Characterized Methanolic and Ethyl Acetate Crude Extracts of A.	

<i>lavus</i>	89
4.5 Secondary metabolites of <i>Aspergillus flavus</i>	93
4.5.1 Methyl (E)-octadec-3-enoate (4.1)	93
4.5.2 Methyl (E)-tetracos-3-enoate (4.2).....	100
4.5.3 Methyl 3-hydroxynonadecanoate (4.3).....	108
4.5.4 (E)-octadec-3-enoic acid (4.4)	114
4.5.5 Methyl (E)-3-hydroxypentadec-5-enoate (4.5)	121
4.5.6 Flavulactone A (4.6).....	128
4.5.7 Flavulactone B (4.7)	134
4.5.8 Flavulactone C (4.8).....	140
4.5.9 Methyl (E)-5-hydroxynonadec-9-enoate (4.9)	146
4.5.10 Flavulactone D (4.10).....	151
4.5.11 Flavulactone E (4.11)	157
CHAPTER FIVE	160
DISCUSSION.....	160
5.1 Diversity of Mangrove Fungal Endophytes	160
5.1.1 Host Recurrence of Mangrove Endophytic Fungi	162
5.1.2 Spatial Heterogeneity of Mangrove Endophytic Fungi.....	163
5.2 Biological Characteristics of Mangrove Fungal endophytes of Coastal Kenya	
5.4 Mangrove Fungal Endophytes of Coastal Kenya.....	166
5.5 Secondary Metabolites from Crude Extracts of <i>Aspergillus flavus</i>	166
5.5.1 Secondary Metabolites from <i>Aspergillus flavus</i>	167
CHAPTER SIX.....	172
CONCLUSIONS AND RECOMMENDATIONS.....	172
6.1 Conclusions	172
6.2 Recommendations.....	173
REFERENCE.....	175

LIST OF APPENDICES.....	196
Appendix I: TUM ERC Certificate	196
Appendix II. Manuscript 1	197
Appendix III: Manuscript 2.....	198
Appendix IV: GC-MS fragmentation for 4.1- 4.8	199

LIST OF FIGURES

Figure 2.1: Schematic diagram of the reaction occurring in polyketide synthases (PKSs); ACP, acyl carrier protein; AT, acyltransferase; KS, ketosynthase; KR, ketoreductase; DH, dehydratase; ER, enoylreductase (<i>Adopted from Risdian et al., 2019</i>)	23
Figure 2.2. The fungal families (<i>Source, https://www.chegg.com 30/3/2022</i>).....	30
Figure 3.1 A map showing the Mangrove forests of Gazi Bay, Tudor and Mida creeks along Kenya coast.....	43
Figure 3.2 A section of degraded mangroves forest of Gazi Bay Kwale County..	44
Figure 3.3 Schematic diagram of isolation, determination of bioactivity, phylogenetic analysis and characterization of mangrove.....	46
Figure 3.4 A schematic diagram of isolation of fungal isolates from selected mangroves of coastal Kenya.....	49
Figure 3.5 A schematic diagram of small-scale fermentation and extraction of Mangrove fungal isolates from selected mangrove species of Coastal Kenya	52
Figure 3.6 Bioassay and preliminary GC-MS analysis of secondary metabolites.....	54
Figure 4.1. A section of the culturally identified mangrove fungal endophytes from the twig cuttings of 4 mangrove species of coastal Kenya (a) <i>A. marina</i> (b) <i>C. tagal</i> (c) <i>R. mucronata</i> (d) <i>S. alba</i>	68
Figure 4.2. Morphologically identified mangrove endophytic fungal genera from selected mangrove species of coastal Kenya; (h) <i>Aspergillus</i> ; (i) <i>Penicillium</i> ; (j)	

<i>Fusarium</i> ; (k) <i>Cephalosporium</i> ; (l) <i>Blastomyces</i>	69
Figure 4.3. Colony morphology of <i>Aspergillus</i> : A) front view; and B) Microscopic features of <i>Aspergillus</i> (x100)	71
Figure 4.4. Colony morphology of <i>Penicillium</i> : A) front view; and B) Microscopic features of <i>Penicillium</i> (x100)	71
Figure 4.5. Colony morphology of <i>Fusarium</i> : A) front view; and B) Microscopic features of <i>Fusarium</i> (x100).....	72
Figure 4.6.Colony morphology of <i>Cephalosporium</i> : A) front view; and B) Microscopic features of <i>Cephalosporium</i> (x100).....	72
Figure 4.7. Colony morphology of <i>Blastomyces</i> : A) front view; and B) Microscopic features of <i>Blastomyces</i> (x100).....	73
Figure 4.8.spatial distribution of mangrove endophytic fungal genera in (e) Tudor Creek, (f) Gazi Bay and (g) Mida Creek	74
Figure 4.9. Colonization of mangrove fungal endophytes sampled along the Kenya coast.....	75
Figure 4.11. Mean inhibition diameters of ethyl acetate crude extracts against <i>Staphylococcus aureus</i> (SA) and <i>Escherichia coli</i> (EC)	81
Figure 4.12.PCR Amplicons (approximately 500 base pairs) of its region of endophytic fungi isolated from selected mangrove species of Coastal Kenya.....	83
Figure 4.13. Phylogenetic tree of 9 unique fungal isolates of Coastal Kenya rooted from <i>Fusarium oxysporum</i>	85
Figure 4.14.(A) A brown gummy ethyl acetate crude extract (B) a brown waxy solid of ethyl acetate crude extract	88

Figure 4.15. Silica gel column chromatography fractions of the methanolic crude extracts of <i>Aspergillus flavus</i>	91
Figure 4.16. Silica gel column chromatography fractions of the ethyl acetate crude extracts of <i>Aspergillus flavus</i>	92
Figure 4.17. Important HMBC correlation diagram of methyl (E)-octadec-3-enoate (4.1).....	94
Figure 4.18. MS fragmentation for methyl (E)-octadec-3-enoate (4.1).....	95
Figure 4.19. ¹ H NMR spectrum of methyl (E)-octadec-3-enoate (4.1).....	97
Figure 4.20. ¹³ C NMR and DEPT NMR spectra of methyl (E)-octadec-3-enoate (4.1)	98
Figure 4.21. H COSY plot for methyl (E)-octadec-3-enoate (4.1)	99
Figure 4.22. HSQC plot for methyl (E)-octadec-3-enoate (4.1).....	99
Figure 4.23. HMBC plot for methyl (E)-octadec-3-enoate (4.1).....	100
Figure 4.24. Important HMBC Correlation diagram of methyl (E)-tetracos-3-enoate (4.2).....	102
Figure 4.25. MS fragmentation of methyl (E)-tetracos-3-enoate (4.2)	102
Figure 4.26. ¹ H NMR spectrum of methyl (E)-tetracos-3-enoate (4.2)	104
Figure 4.27. ¹³ C NMR and DEPT NMR spectra of methyl (E)-tetracos-3-enoate (4.2)....	105
Figure 4.28. ¹ H COSY plot for methyl (E)-tetracos-3-enoate (4.2).....	106
Figure 4.29. HMBC plot for methyl (E)-tetracos-3-enoate (4.2).....	107
Figure 4.30. Important HMBC Correlation for methyl 3-hydroxynonadecanoate (4.3) .	109
Figure 4.31. MS fragmentation for methyl 3-hydroxyoctadecanoate (4.3)	109
Figure 4.32. ¹ H NMR spectrum of methyl 3-hydroxyoctadecanoate (4.3).....	111

Figure 4.33. ^{13}C NMR and DEPT NMR spectral data of methyl 3-hydroxyoctadecanoate (4.3)	112
Figure 4.34. ^1H COSY plot for methyl 3-hydroxyoctadecanoate (4.3)	113
Figure 4.35. HSQC plot for methyl 5-hydroxyoctadecanoate (4.3)	113
Figure 4.36. HMBC plot for methyl 3-hydroxyoctadecanoate (4.3)	114
Figure 4.37: HMBC Correlation diagram of (E)-octadec-3-enoic acid (4.4)	115
Figure 4.38. MS fragmentation for (E)-octadec-3-enoic acid (4.4)	116
Figure 4.39. ^1H NMR spectrum of (E)-octadec-3-enoic acid (4, 4)	118
Figure 4.40. ^{13}C NMR and DEPT NMR spectra for (E)-octadec-3-enoic acid (4.4)	119
Figure 4.41. ^1H COSY plot for (E)-octadec-3-enoic acid (4.4)	120
Figure 4.42. HSQC plot for (E)-octadec-4-enoic acid (4.4)	120
Figure 4.43. HMBC plot for (E)-octadec-3-enoic acid (4.4)	121
Figure 4.44. Important HMBC Correlation of methyl (E)-3-hydroxypentadec-5-enoate (4.5)	122
Figure 4.45. MS fragmentation for methyl (E)-3-hydroxypentadec-5-enoate (4.5).	123
Figure 4.46. ^1H NMR spectrum of methyl (E)-3-hydroxypentadec-5-enoate (4.5)	125
Figure 4.47. ^{13}C NMR and DEPT NMR spectra of Methyl (E)-3-hydroxypentadec-5-enoate (4.5)	126
Figure 4.48. ^1H COSY plot for methyl (E)-3-hydroxypentadec-5-enoate (4.5)	127
Figure 4.49. HSQC plot for methyl (E)-3-hydroxypentadec-5-enoate (4.5)	127
Figure 4.50. HMBC plot for methyl (E)-3-hydroxypentadec-5-enoate (4.5)	128
Figure 4.51. Important HMBC correlations for Flavulactone A (4.6)	129

Figure 4.52. ¹ H NMR for Flavulactone A (4.6).....	131
Figure 4.53. ¹³ C NMR and DEPT NMR for Flavulactone A (4.6).....	132
Figure 4.54. ¹ H COSY plot for Flavulactone A (4.6).....	133
Figure 4.55. HSQC plot for Flavulactone A (4.6).....	133
Figure 4.56. HMBC plot for Flavulactone A (4.6).....	134
Figure 4.57. Important HMBC correlations of Flavulactone B (4.7).....	135
Figure 4.58. ¹ H NMR spectrum of Flavulactone B (4.7).....	137
Figure 4.59. ¹³ C NMR and DEPT NMR spectra of Flavulactone B (4.7).....	138
Figure 4.60. ¹ H COSY plot for Flavulactone B (4.7).....	139
Figure 4.61. HSQC plot for compound Flavulactone B (4.7).....	139
Figure 4.62. ¹ HMBC plot for Flavulactone B (4.7).....	140
Figure 4.63. Important HMBC correlations of Flavulactone C (4.8).....	141
Figure 4.64. ¹ H NMR for Flavulactone C (4.8).....	143
Figure 4.65. ¹³ C NMR and DEPT NMR spectral data of Flavulactone C (4.8).....	144
Figure 4.66. ¹ H COSY plot for Flavulactone C (4.8).....	145
Figure 4.67. HSQC plot for Flavulactone C (4.8).....	145
Figure 4.68. HMBC plot for Flavulactone C (4.8).....	146
Figure 4.69. HMBC correlations of Methyl (E)-5-hydroxynonadec-9-enoate (4.9).....	147
Figure 4.70. ¹ H NMR for methyl (E)-5-hydroxynonadec-9-enoate (4.9).....	149
Figure 4.71. ¹ H COSY of methyl (E)-5-hydroxynonadec-9-enoate (4.9).....	150
Figure 4.72: HSQC correlations of methyl (E)-5-hydroxynonadec-9-enoate (4.9).....	150
Figure 4.73. HMBC correlations of methyl (E)-5-hydroxynonadec-9-enoate (4.9).....	151

Figure 4.74: HMBC correlations for Flavulactone D (4.10).....	152
Figure 4.75: ¹ H NMR for Flavulactone D (4.10).....	154
Figure 4.76: ¹³ C NMR of Flavulactone D (4.10).....	155
Figure 4.77: ¹ H, H- COSY NMR for Flavulactone D (4.10)	156
Figure 4.78: HSQC correlations for Flavulactone D (4.10)	156
Figure 4.79. HMBC correlations of Flavulactone D (4.10).....	157
Figure 4.80. HMBC correlations of methyl (E)-5-(4-hydroxy-2-((E)-3-hydroxytetradec- 1-en-1-yl)-6-oxotetrahydro-2H-pyran-3-yl) pent-3-enoate (4.11).....	158
Figure 5.1: Biosynthetic pathway for methyl-3-hydroxyoctadecanoate (4.3).....	168
Figure 5.2 Joint biosynthetic pathway of compound 4.1 to 4.5.....	169
Figure 5.3: Folding and cyclization of 5-hydroxyhexadecan-2-one in flavulactone B formation.....	170
Figure 5.4: Suggested biogenetic pathway for compounds 4.6, 4.7 and 4.8	171

LIST OF TABLES

Table 2.1: Selected mangrove species of Kenya and their uses (<i>Source</i> Bandaranayake, 1998)	Error! Bookmark not defined. 9
Table 4.1: Morphological characteristics of fungal mangrove endophytes isolated from selected study sites in coastal Kenya	70
Table 4.2. Colonization and isolation rates of mangrove fungal endophytes sampled along the Kenya coast.....	75
Table 4.3. Antibacterial activities of mangrove fungal isolates against selected pathogens from coastal Kenya.....	78
Table 4.4. Mean inhibition diameters for <i>S. aureus</i> and <i>E. coli</i> at different concentrations.....	80
Table 4.5. Minimum inhibition concentration in mg/ml of ethyl acetate extract of <i>Aspergillus flavus</i> from Coastal Kenya.....	82
Table 4.6: Concentration and Purity of DNA Extract.....	83
Table 4.7. Mangrove Fungal Isolates Identified by Molecular Techniques.....	86
4.8. Results of GC-MS analysis of ethyl acetate crude extracts of <i>Aspergillus flavus</i>	89
Table 4.9. ¹ H NMR and ¹³ C NMR(400 MHz) spectral data for methyl (E)-octadec-	96
Table 4.10. ¹ H NMR and ¹³ C NMR(400 MHz) spectral data for methyl 3-hydroxyoctadecanoate (4.2)	103
Table 4.11. ¹ H NMR and ¹³ C NMR(400 MHz) spectral data for (E)-octadec-3-enoic acid(4.3).....	110
Table 4.12. ¹ H NMR and ¹³ C NMR(400 MHz) spectral data for methyl (E)-3-h.....	117
Table 4.13. ¹ H NMR and ¹³ C NMR(400 MHz) spectroscopic data for Flavulactone A (4.6).....	124
Table 4.14: ¹ H NMR and ¹³ C NMR(400 MHz) spectra for Flavulactone B (4.7)	130
Table 4.15: ¹ H NMR and ¹³ C NMR(400 MHz) spectral data of Flavulactone C (4.8)	136
Table 4.16. ¹ H NMR and ¹³ C NMR(400 MHz) spectroscopic data for methyl (E)-5-hydroxynonadec-9-enoate (4.9)	142

Table 4.17 ^1H NMR and ^{13}C NMR(400 MHz) spectroscopic data of Flavulactone D (4.10)

Table 4.18. ^1H NMR and ^{13}C NMR(400 MHz) spectroscopic data formethyl (E)-5-(4-

hydroxy-..... 153

Table 4.19. ^1H NMR and ^{13}C NMR (400 MHz) spectroscopic data formethyl (E)-5-(4-

hydroxy-2-(E)-3-hydroxytetradec-1-en-1-yl)-6-oxotetrahydro-2H-pyran-3-yl) pent-3-

enoate(4.11).....159

LIST OF ABBREVIATIONS

MFEs	Mangrove fungal endophytes
μ	Micro
μg	Microgram
μL	Microliter
¹³ C NMR	Carbon Nuclear Magnetic Resonance
¹³ C	Carbon thirteen isotope
1D	One dimensional NMR
¹ H NMR	Proton Nuclear Magnetic Resonance
¹ H	Proton
2D	Two-dimensional NMR
3α-HSD	3α-hydroxysteroid dehydrogenase
ACD	Advance chemistry development
AF ₁₋₁₆	<i>Aspergillus flavus</i> 1-16
AMR	Antimicrobial resistance
ANOVA	Analysis of variance
ATCC	American type culture collection
BLAST	Basic alignment search tool
CFU	Colony Forming Units
CO ₂	Carbon Dioxide
COSY	Correlation Spectroscopy
COVID	Corona Virus Disease
COX	Chemiluminescent Cyclooxygenase
CTAB	Cetyl Trimethylammonium Bromide
DCM	Dichloromethane
DEPT	Distortionless Enhancement by Polarization Transfer
DMSO	Dimethyl Sulphoxide
DNA	Deoxyribonucleic Acid

EAE	Ethyl Acetate Extract
FAME	Fatty acid methyl ester
EtOH	Ethanol
g	gram
GC-MS	Gas chromatography-Mass spectrometry
H1N1	Influenza virus
Ha	Hectare
HMBC	Heteronuclear Multiple Bond Correlation
HSQC	Heteronuclear Simple Quantum Correlation
HUS	Hemolytic uremic syndrome
IDs	Infectious Diseases
IR	Infrared
ITS	Internal transcribed spacer
IZDs	Inhibition zone diameters
KMFRI	Kenya Marine and Fisheries Research Institute
MDG	Millennium development goal
MDR	Multi Drug Resistance
ME	Methanolic extract
MEGA	Molecular evolutionary genetics analysis
MeOH	Methanol
MHA	Muller Hinton Agar
MIC	Minimum inhibition concentration
mL	Milli Litre
MS	Mass Spectrometry
MUSCLE	Multi sequence Alignment
NA	Nutrient Agar
NACOSTI	National Commission for Science, Technology and Innovation
NaHCO ₃	Sodium hydrogen sulphate
NC	Non-Clavicipitaceous

NCBI	National center for biological information
NMR	Nuclear magnetic resonance
NRF	National Research Fund
°C	Degrees Celsius
PCR	Polymerase chain reaction
PDA	Potato dextrose Agar
PDB	Potato dextrose broth
SARS	Severe acute respiratory syndrome
SD	Standard deviation
SE	Standard error
TLC	Thin layer chromatography
TUM	Technical University of Mombasa
TUM ERC	Technical University of Mombasa Ethical Review Committee
UV	Ultraviolet
WHO	World Health Organization
WIOMSA	Western Indian Ocean Marine Science Association
ZIDs	Zone Inhibition Diameters
SDG	Sustainable development goals
MDH	Malate dehydrogenase
TMS	Tetramethyl silane
MRSA	Methicillin-resistant <i>Staphylococcus aureus</i>
PKS	Polyketide Synthase
MHB	Mycorrhiza helper bacteria

ABSTRACT

Due to the global spread of resistant bacteria and fungi, antibiotics are no longer as effective as they once were, and microbial illnesses are once more a threat to human life. This situation is offering a continuous opportunity for research of alternative novel bioactive molecules to address the problem. In this study we isolated and characterized bioactive compounds from selected endophytic fungi originating from common mangroves of the Kenya coastline namely; *Avicennia marina*, *Ceriops tagal*, *Rhizophora mucronata* and *Sonneratia alba* against *Staphylococcus aureus* and *Escherichia coli* that could potentially be used for drugs development. Isolation and purification using potato dextrose agar (PDA), potato dextrose broth (PDB) gave 19 mangrove fungal endophytes (MFEs). Morphological identification resulted in 18 MFEs belonging to 5 fungal genera namely; *Aspergillus*, *Penicillium*, *Fusarium*, *Cephalosporium* and *Blastomyces*. Molecular identification gave 9 successfully characterized species belonging to the genus *Aspergillus* namely; *Aspergillus flavus*, *Aspergillus niger*, *Aspergillus nomius*, *Aspergillus tubingensis*, *Aspergillus oryzae*, *Aspergillus awamori*, *Aspergillus aculeatus*, *Aspergillus brunneoviolaceus* and *Aspergillus welwitchiae*. Ethyl acetate extract (EAE) and methanolic extract (ME) were produced through fermentation and extraction with ethyl acetate and ethyl acetate in 10% methanol. The extracts contained alkaloids, phenols, steroids, tannins, and terpenoids, according to preliminary phytochemical screening. *Aspergillus flavus* ethyl acetate extracts showed maximum activity when tested for susceptibility via disk diffusion on Muller Hinton Agar, with inhibition zone diameters (IZDs) Standard deviation (SD) of (17.1 ± 2.45) and (15.9 ± 2.45) against *S. aureus* and *E. coli*, respectively. *Aspergillus flavus* methanolic extracts had a lower inhibition activity (9.2 ± 0.75) and no action against *E. coli* at the investigated doses as compared to the positive control. The minimum inhibitory concentration of *A. flavus* crude extract against *S. aureus* and *E. coli* was (0.82 ± 0.05) and (0.91 ± 0.05) mg/ml, respectively. Results of one-way ANOVA test indicated no significant difference in the inhibition between the two test pathogens ($p > 0.05$). Results of Gas Chromatography coupled with Mass Spectrometry (GC-MS) analysis of ethyl acetate crude extracts of *A. flavus* revealed four known active compounds namely; Lactic acid, Isopropyl alcohol, Semi carbazone and Corydaldine. All of them were active against a broad spectrum of pathogens including *S. aureus* and *E. coli* in this study. Fractionation using Silica gel (60-120) mesh column chromatography of the antimicrobial *A. flavus* methanolic and ethyl acetate extract, gave 11 compounds which when characterized by TLC resulted in pure compounds with different RF values. The isolates were established to be the new δ -lactones Flavulactone B, Flavulactone C, Flavulactone D, Flavulactone E, and Flavulactone F which were all new compounds alongside the known 6-tridecyloxan-2-one now named Flavulactone A. The other oxy-compounds were the new fatty acid methyl esters; methyl (E) - octadec-3-enoate, methyl (E)- tetracos-3-enoate and Methyl (E)-3-hydroxypentadec-5-enoate, methyl (E)-5-hydroxynonadec-9-enoate and the known methyl 3-hydroxynonadecanoate and (E)-octadec-3-enoic acid. The study confirms that

bioactive metabolites indeed reside in endophytic fungi inhabiting selected mangroves from coastal Kenya.